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# Praesalide E, a new phenolic compound from the lichen Parmotrema praesorediosum (nyl.) Hale

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#### ABSTRACT

Introduction: Parmotrema praesorediosum (Nyl.) Hale is widely distributed in the south of Vietnam. To contribute to finding new compositions as well as increasing the phytochemical knowledge of Parmotrema species from which we could further study to find some new biologically potential compounds, we have done systematic research on Parmotrema praesorediosum growing in Vietnam, and we have been reported twelve novel phenolic compounds. This paper reported the isolation and structural determination of two compounds as the result of the continuous study on the chloroform extract of Parmotrema praesorediosum. Method: The crude extract was obtained by the maceration method of dried power of Parmotrema praesorediosum lichen in methanol. This extract was then separated by the solid-phase extraction and eluted with petroleum ether, chloroform, ethyl acetate, acetone, and methanol in turn to give the corresponding extracts. Two compounds were isolated from chloroform extract by silica gel column chromatography. Their chemical structures were determined by the NMR and HR-ESI-MS data analysis. Moreover, the chemical structure of **1** was also deduced from its methylated product. The cytotoxicity of compound **2** against HeLa, NCI-H460 and, MCF-7 cell lines was done using the Sulforhodamine B method. Results: Two isolated compounds, consisting of praesalide E (1) and usenamin A (2), and one methylated product from 1 were identified. Compound 2 showed no toxicity against all three tested cell lines at the concentration of 100  $\mu$  g/mL. **Conclusion:** To the best of our knowledge, **1** was a new compound, and compound 2 was known to present in *Parmotrema* genus for the first time. Key words: Parmotrema praesorediosum, lichen, cytotoxicity, praesalide.

# **INTRODUCTION**

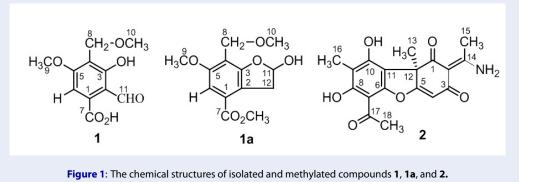
The studies of drugs originated from natural products or based on natural product models have been studied a lot because of their high effectiveness and few side effects. Lichens, symbiotic associations between fungi and algae, contain unique substances such as depsides, depsidones, diphenyl ethers... Therefore, lichens are attractive for pharmacological and chemical studies<sup>1-3</sup>, whereas lichens habited in Vietnam, a tropical region, have not much been chemically studied. To contribute to finding new compositions as well as increasing the phytochemical knowledge of Parmotrema spieces, we have done systematic research on lichen substances from the Vietnamese flora and reported twelve novel phenolic compounds, including eight diphenyl ethers, three phthalide derivatives, and one monoaromatic compound<sup>4-8</sup> from Parmotrema praesorediosum (Nyl.) Hale, which is widely distributed in the south of Vietnam. Herein, we continuously report the isolation and structural elucidation of a new phenolic praesalide E (1) and usenamin A (2) and a cytotoxic ability against HeLa, NCI-H460, and MCF-7 cell lines of compound 2 as well. Their chemical structures (Figure 1) were unambiguously determined by analyzing 1D and 2D NMR and highresolution ESI mass spectroscopic data and comparing their NMR data with the ones published in the literature.

## **MATERIALS AND METHODS**

## **General experimental procedures**

Column chromatography was performed on silica gel (Merck) (230-400 Mesh). Thin-layer chromatography (TLC) and preparative TLC were performed on silica gel GF<sub>254</sub> (Merck), visualized by vanillin, followed by heating. NMR spectra were acquired on Bruker 500 Avance III at 500 MHz for <sup>1</sup>H-NMR and 125 MHz for <sup>13</sup>C-NMR. The HR-ESI-MS spectra were recorded on a Bruker microOTOF Q-II in the Center Analysis Laboratory of the University of Science, Vietnam National University (VNU)– Ho Chi Minh City. The optical rotations were measured on Krüss (German) digital polarimeter. All solvents were used for extraction and purification from ChemSol manufacturer and redistillation.

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#### **Plant material**

The lichen *Parmotrema praesorediosum* (Nyl.) Hale was collected at Nam Cat Tien National Forest Reserve and Intermediate Zones, Nam Cat Tien Village, Tan Phu District, Dong Nai Province, Vietnam. The scientific name of the lichen was authenticated by Dr. Vo Thi Phi Giao, Faculty of Biology, University of Science, VNU HCM. A voucher specimen (No US-B020) was deposited in the Herbarium of the Department of Organic Chemistry, Faculty of Chemistry, University of Science, VNU HCM.

#### **Extraction and isolation**

5.0 Kg of the fresh lichen thalli were cleaned under running tap water and air-dried. The ground powder (3.0 kg) was exhaustedly extracted with methanol by the method of maceration. The filtrated solution was evaporated at reduced pressure. While the methanolic solution was evaporated, a precipitate appeared and was filtered off, and then the solution was continued to evaporate to dryness. The resulting was the precipitate (9.0 g) and the crude methanolic residue (450.0 g). The methanolic residue (450.0 g) was subjected to silica gel solid-phase extraction and eluted consecutively with petroleum ether, chloroform, ethyl acetate, acetone, and methanol in turn at room temperature to afford petroleum ether extract (E1 25.0 g), petroleum ether extract (E2 15.0 g), chloroform extract (C, 105.0 g), ethyl acetate extract (EA, 50.0 g), acetone extract (AC, 45.0 g) and methanol extract (M, 37.0 g).

The chloroform extract was subjected to silica gel column chromatography, eluted with the solvent system of petroleum ether – ethyl acetate (0-100% of ethyl acetate) to obtain twenty-three fractions from C1 to C23. Fraction C16 (4.2 g) was rechromatographed, eluted with petroleum ether-chloroform (5:5) to give the compound **2** (15.0 mg). Fraction C20 (23.9 g) was repeatedly subjected to silica gel column chromatography, eluted with *n*-hexane–diethyl ether (2:8) and chloroform-methanol (98:2) to afford compound **1** (15.7 mg).

## Methylation of 1

TMS-CH<sub>2</sub>N<sub>2</sub> in *n*-hexane were added to a solution of **1** (12.0 mg) in Et<sub>2</sub>O (1 mL) and MeOH (0.5 mL). The mixture was stirred at room temperature for 1 hour and 15 mins. After termination by diluted acetic acid in MeOH, the reaction mixture was concentrated *in vacuo*, and the residue was purified by preparative TLC (*n*-hexane-Et<sub>2</sub>O, 2:8) to yield compound **1a** (4.1 mg).

## **Cytotoxicity inhibitory activities**

The testing of cytotoxic activities against the MCF-7 (breast cancer cell line), HeLa (cervical cancer cell line), and NCI-H460 (lung cancer cell line) was done using the Sulforhodamine B method (SBR assay), described by Skehan with camptothecin as a positive control<sup>9</sup>. This experiment was described in more detail by Nguyen *et al.*<sup>10</sup> and was done at the Department of Molecular Biology, Faculty of Biology, University of Science, VNU HCM.

## RESULTS

From the chloroform extract of the lichen *P. praesorediosum* at Nam Cat Tien National Forest Reserve and Intermediate Zones, Dong Nai Province, two compounds, **1** (15.7 mg) and **2** (15.0 mg), were isolated. 4.1 mg of **1a** was synthesized and purified from 12.0 mg of **1**. Their physical properties and spectroscopic data were performed as following and in **Table 1**.

**Praesalide E (1):** Yellow solid. HR-ESI-MS: m/z 241.0704 [M+H]<sup>+</sup> and m/z 263.0523 [M+Na]<sup>+</sup>. <sup>1</sup>H and <sup>13</sup>C-NMR (DMSO- $d_6$ ) data: see **Table 1**. Selected HMBC correlations: see **Figure 2**.

**Compound 1a:** Yellow solid.  $[\alpha]_D^{21}$  +6.6 (*c* 0.85, CHCl<sub>3</sub>). IR(KBr)  $v_{max}$  cm<sup>-1</sup>: 3477, 1725, 1626, 1462, 1238. HR-ESI-MS: *m/z* 269.1022 [M+H]<sup>+</sup> and *m/z* 

291.0840 [M+Na]<sup>+</sup>. <sup>1</sup>H and <sup>13</sup>C-NMR (CDCl<sub>3</sub>) data: see **Table 1**. Selected COSY, HMBC and NOESY correlations: see **Figure 2**.

(+)-(12*R*)-Usenamin A (2): Yellow solid,  $[\alpha]_D^{23}$ +852 (*c* 0.001, MeOH); HR-ESI-MS: *m/z* 366.0938 [M+Na]<sup>+</sup>. The <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta_H$  13.35 (1H, s, 8-OH), 11.76 (1H, s, 10-OH), 5.82 (1H, s, H-4), 2.68 (3H, s, H-18), 2.62 (3H, s, H-15), 2.09 (3H, s, H-16), 1.70 (3H, s, H-13). <sup>13</sup>C-NMR (CDCl<sub>3</sub>):  $\delta_C$  199.0 (C-1), 102.4 (C-2), 190.8 (C-3), 102.7 (C-4), 174.9 (C-5), 155.9 (C-6), 101.6 (C-7), 163.7 (C-8), 108.4 (C-9), 158.3 (C-10), 105.0 (C-11), 57.4 (C-12), 31.9 (C-13), 175.5 (C-14), 26.6 (C-15), 7.6 (C-16), 200.8 (C-17), 31.4 (C-18). Selected HMBC correlations: see **Figure 2**.

Compound **2** was tested the cytotoxic activity against three cell lines HeLa (human epithelial carcinoma), NCI-H460 (human lung cancer), and MCF-7 (human breast cancer) at the concentration of 100  $\mu$ g/mL. The percentages of inhibition of cell growth (I %) against HeLa, NCI-H460, and MCF-7 were 18.9  $\pm$  3.4, 6.0  $\pm$  0.1, and 3.7  $\pm$  1.2, respectively, which showed very low cytotoxicity against all tested cell lines at the concentration of 100  $\mu$ g/mL.

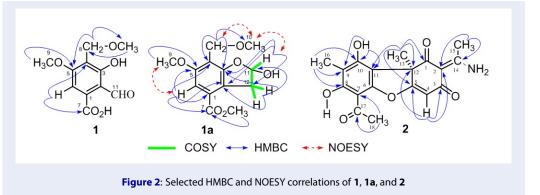
## DISCUSSION

Compound 1 was isolated as a yellow solid. Its molecular formula was determined as C11H12O6 based on the protonated ion peak at m/z 241.0704 [M+H]<sup>+</sup>, (Calcd. for C<sub>11</sub>H<sub>12</sub>O<sub>6</sub>+H, 241.0712) and the sodiated adduct at m/z 263.0523 [M+Na]<sup>+</sup>, (calcd. for  $C_{11}H_{12}O_6$ +Na, 263.0531). The <sup>1</sup>H-NMR spectrum of 1 showed one chelated hydroxyl proton (1H,  $\delta_H$ 12.47, s), one aldehydic proton (1H,  $\delta_H$  10.42, s), only one aromatic methine proton (1H,  $\delta_H$  6.86, s) of a pentasubstituted benzene ring. At a higher magnetic field, the presence of methylene protons at  $\delta_H$ 4.40 (2H, s, H-8) showing HMBC cross-peaks to methoxy carbon at  $\delta_C$  57.3 (C-10) constructed the -CH<sub>2</sub>-O-CH<sub>3</sub> group in its chemical structure. The rest methoxy proton at  $\delta_H$  3.87 (3H, s, H-9) and the proton H-8 showed HMBC correlations to the same oxygenated aromatic carbon at  $\delta_C$  162.9 (C-5), which suggested two adjacent positions on benzene ring of these two groups. From the molecular formula of C11H12O6, its degree of unsaturation were calculated to be 6, including 4 of the benzene ring, 1 of aldehyde group, and the rest of a carboxyl carbon which was determined via the observation of a signal at  $\delta_C$  168.4 (C-7) on the <sup>13</sup>C-NMR spectrum. All above analyses, as well as 12 hydrogen atoms in its molecular, inferred

that **1** possessed one benzene ring with five substitutions including -OH, -CHO, -OCH<sub>3</sub>, -CH<sub>2</sub>OCH<sub>3</sub>, -COOH. However, the <sup>13</sup>C-NMR spectrum of **1** displayed only six carbon signals, and the HMBC spectrum did not show enough correlation to determine its chemical structure unambiguously. Therefore, it could be deduced via its methylated product.

Methylation of 1 by diazomethane afforded 1a to be as a yellow solid. The HR-ESI-MS spectrum of 1a displayed the pseudo molecular ion peak at m/z 269.1022  $[M+H]^+$ , (Calcd. for C<sub>13</sub>H<sub>16</sub>O<sub>6</sub>+H, 269.1025) and m/z 291.0841 [M+Na]<sup>+</sup>, (calcd. for C<sub>13</sub>H<sub>16</sub>O<sub>6</sub>+Na, 291.0845), which approved its molecular formula of C13H16O6. The combination of 1D and 2D-NMR data analysis showed that 1a possessed similar structure to 1, including a pentasubstituted benzene ring [5 quaternary aromatic carbons ( $\delta_C$  158.8, 158.5, 126.5, 120.4, 113.1) and one aromatic methine ( $\delta_H$  7.09, s;  $\delta_C$  104.1)], a methoxy group [C-9 ( $\delta_H$  3.87, s;  $\delta_C$ 56.2)], a methoxymethylene group [C-8 ( $\delta_H$  4.52, d, J = 10.5 Hz, 4.56, d, J = 10.5 Hz;  $\delta_C$  63.1), C-10 ( $\delta_H$ 3.37, s;  $\delta_C$  58.2)], a carboxyl group [C-7 ( $\delta_C$  166.6)]. The positions of the carboxyl, methoxymethylene, and methoxy groups were determined at C-1, C-4, C-5 of benzene ring, respectively, via HMBC correlations as shown in Figure 2. The differences between 1 and 1a were the absence of the chelated hydroxyl proton and the aldehydic proton in 1a. Meanwhile, it displayed one more methoxy group [C-13 ( $\delta_H$  3.90, s;  $\delta_C$ 52.1)], one more methylene group [C-12 ( $\delta_H$  3.37, dd,  $J = 18.0, 2.5 \text{ Hz and } 3.57, \text{ dd}, J = 18.0, 6.5 \text{ Hz}; \delta_C 38.9)$ ] and one more hemiacetalic group [C-11 ( $\delta_H$  6.15, dd, J = 6.5, 2.5 Hz;  $\delta_C 101.8$ ). The methoxy group attached to carboxyl carbon (C-7) was confirmed by HMBC correlation of this methoxy proton and C-7. The COSY cross-peak between the acetalic proton and the methylene protons as well as HMBC correlations of these protons to the same aromatic carbons at  $\delta_C$ 120.4 (C-2) and 158.5 (C-3) suggested the appearance of a benzofuran skeleton. The NOESY interaction from the acetalic proton to the methoxy protons of the methoxymethylene group suggested the positions of the acetal carbon and the methylene carbon in the benzofuran moiety (Figure 2). Complete analysis of the 2D-NMR data for 1a resulted in its formulation, as shown in Figure 1.

From the chemical structure of 1a, the one of 1 was deduced via a proposed retrosynthesis as presented in Scheme 1. The conversion of the carboxylic acid group (C-7) in 1 to a methyl ester in 1a by diazomethane is a simple reaction. The formation of the furan ring could be explained as follows: firstly, diazomethane reacted with the aldehyde group of 1 to



form a homologated aldehyde in a modification of the Buchner-Curtius-Schlotterbeck reaction<sup>11</sup>. The next reaction has been extended to form a hemiacetal by a nucleophilic addition between the pair of free electrons on the phenolic hydroxyl group and the aldehyde group. Furthermore, re-analyses of HMBC spectrum for 1 revealed that H-6 ( $\delta_H$  6.86) correlated with a carbon signal at  $\delta_C$  113.3 (C-4) and the methoxy group H<sub>3</sub>-9 ( $\delta_H$  3.87) as well as the methylene protons H<sub>2</sub>-8 ( $\delta_H$  4.40) correlated with an oxygenated aromatic carbon signal at  $\delta_C$  162.9 (C-5), confirming the presence of aromatic carbons C-4 and C-5 (Figure 2). These results suggested the chemical structure of 1 as 2-formyl-3-hydroxy-5-methoxy-4-methoxymethylbenzoic acid, namely praesalide E. Compound 2 was obtained as a vellow solid. Its molecular formula was determined as C18H17O6N through its pseudo molecular ion peak at m/z 366.0938 [M+Na]<sup>+</sup> (calcd. 366.0954 for C<sub>18</sub>H<sub>17</sub>O<sub>6</sub>N+Na) in the HR-ESI-MS spectrum. The <sup>1</sup>H-NMR spectrum exhibited signals of two chelated hydroxyl groups at  $\delta_H$  11.76 (1H, s, 10-OH), and 13.35 (1H, s, 8-OH); four methyl groups at  $\delta_H$  1.70 (3H, s, H-13), 2.09 (3H, s, H-16), 2.62 (3H, s, H-15), and 2.68 (3H, s, H-18); and an aromatic proton at  $\delta_H$  5.82 (1H, s, H-4). These protons enabled the identification of 2 possessing the 9bH-dibenzofurandione moiety. The <sup>13</sup>C-NMR spectrum showed 18 carbons including three keto carbonyl carbons at  $\delta_C$  199.0 (C-1), 190.8 (C-3), and 200.8 (C-17); nine olefinic carbons at  $\delta_C$  101.6 (C-7), 102.6 (C-4), 102.4 (C-2), 105.0 (C-11), 108.4 (C-9), 158.3 (C-10), 155.9 (C-6), 163.7 (C-8), and 174.9 (C-5) among them five carbons were oxygenated; one saturated quaternary carbon at  $\delta_C$  57.4 (C-12); signals of four methyl groups at  $\delta_C$  7.6 (C-16), 26.6 (C-15), 31.4 (C-18), and 31.9 (C-13). The rest carbon signal appearing at  $\delta_C$  175.5 (C-14) was assigned for an enamine group which was demonstrated by the presence of a nitrogen atom

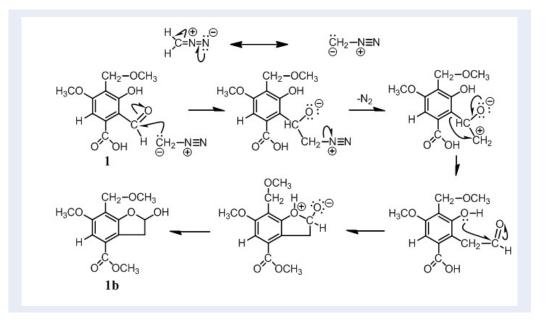
in the molecule and the HSQC and HMBC data analysis. Some selected HMBC correlations of **2** were presented in Figure 2, which approved its plana chemical structure. The absolute configuration of the unique chiral center C-12 was suggested to be *R* due to the biosynthetic aspect that **2** and two similar compounds, usnic acid and isousnic acid possessing 12-*R* configuration<sup>12</sup>, were isolated from the same material and were all dextrorotary. Additionally, the comparison of these spectroscopic data of **2** with those of (+)-(*12R*)-usenamin A in the literature<sup>13</sup> showed good compatibility.

## CONCLUSION

From the continuous phytochemical investigation of the chloroform extract of the lichen *Parmotrema praesorediosum*, two phenolic compounds were isolated, consisting of praesalide E (1) and usenamin A (2). Their chemical structures were established primarily by NMR, MS spectroscopic data, optical rotations analysis as well as deduced from the methylated product. Compound 1 was a new compound, while compound 2 was isolated from *Parmotrema* genus for the first time. Compound 2 had no toxicity against all three tested cell lines, HeLa, NCI-H460, and MCF-7, at the concentration of 100  $\mu$ g/mL.

## **ABBREVIATIONS**

HR-ESI-MS: High resolution-Electrospray ionization-Mass spectrometry <sup>1</sup> H-NMR: Proton nuclear magnetic resonance <sup>13</sup> C-NMR: Carbon-13 nuclear magnetic resonance COSY: Correlation spectroscopy HSQC: Heteronuclear single quantum coherence HMBC: Heteronuclear multiple bond correlation NOESY: Nuclear Overhauser Effect Spectroscopy *s*: singlet *brs*: broad singlet *d*: doublet *dd*: doublet of doublets



Scheme 1: Mechanism for the methylation of 1

No.	1*			1a#		
	$\delta_H$	J (Hz)	$\delta_C$	$\delta_H$	J (Hz)	$\delta_C$
1						126.5
2						120.4
3						158.5
4			113.3			113.1
5			162.9			158.7
6	6.86			7.09		104.1
7			168.4			166.6
8	4.40		61.1	4.56	d (10.5)	63.1
				4.52	d (10.5)	
9	3.87		56.0	3.87		56.2
10	3.21		57.3	3.37		58.2
11	10.42	brs		6.15	dd (6.5, 2.5)	101.8
12				3.38	dd (18.0, 2.5)	38.9
				3.57	dd (18.0, 6.5)	
13				3.90		52.1
3-OH	12.47					

*Note:* \*: dimethylsulfoxide-*d*<sub>6</sub> #: chloroform-*d* 

## **COMPETING INTEREST**

The authors declare no competing financial interest.

## **AUTHORS' CONTRIBUTION**

Huynh B.L.C has contributed to conducting experiments, acquisition of data, and interpretation of data. Nguyen T. A. T., Vo T. N., Pham N. K. T. interpreted NMR and MS data as well as searched the bibliography. Nguyen T. H. T gave final approval of the manuscript to be submitted.

## REFERENCES

- 1. Ahmadjian V, Hale ME. The lichens. Academic Press, New York and London. 1973;.
- Nash III TH. Lichen biology, 2nd Ed. Cambridge University Press, Cambridge, New York. 2008;.
- Huneck S, Yoshimura I. Identification of lichen substances. Springer-Verlag Berlin Heidelberg, New York. 1996;Available from: https://doi.org/10.1007/978-3-642-85243-5.
- Huynh BLC, Le HD, Takenaka Y, Tanahashi T, Nguyen KPP. New phenolic compounds from the lichen Parmotrema praesorediosum (Nyl.) Hale. Magnhetic Resonance Chemistry. 2016; 54:81-87;PMID: 26303251. Available from: https://doi.org/10. 1002/mrc.4316.
- Bui VM, Khong VKL, Huynh BLC, Duong TH, Ha XP, Tanahashi T, Nguyen KPP. A new diphenyl ether from the lichen Parmotrema praesorediosum (Nyl.) Hale Parmeliaceae. Journal of Science and Technology. 2016; 54:77-83;.
- 6. Huynh BLC, Tran TTA, Pham NKT and Nguyen KPP. A new compound from the lichen Parmotrema praesorediosum

(Nyl.) Hale Parmeliaceae. Vietnam Journal of Chemistry. 2017; 55:172-175;.

- Huynh BLC, Bui VM, Pham NKT, Nguyen KPP, Nguyen TP. Three new diphenyl ethers from the lichen Parmotrema praesorediosum (Nyl.) Hale Parmeliaceae. Natural Product Research. DOI 10.1080/14786419.2020.1837818, 2020;Available from: https://doi.org/10.1080/14786419.2020.1837818.
- Huynh BLC, Pham NKT, Nguyen TP. Vinapraesorediosic acids D and E from the lichen Parmotrema praesorediosum (Nyl.) Hale. Phytochemistry Letter. 2021; 41:61-64;Available from: https://doi.org/10.1016/j.phytol.2020.11.001.
- Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D, Warren JT, Bokesch H, Kenney S, Boyd MRJ. New colorimetric cytotoxicity assay for anticancer-drug screening. Journal of National Cancer Institute. 1990; 82:1107-1112; PMID: 2359136. Available from: https://doi.org/10.1093/jnci/82.13.1107.
- Nguyen THT, Pham HVT, Pham NKT, Quach NDP, Pudhom K, Hansen PE, Nguyen KPP. Chemical constituents from Sonneratia ovata Backer and their in vitro cytotoxicity and acetylcholinesterase inhibitory activities. Bioorganic and Medicinal Chemistry Letters. 2015; 25:2366-2371;PMID: 25933595. Available from: https://doi.org/10.1016/j.bmcl.2015.04.017.
- Wang Z. Comprehensive organic name reactions and reagents. Wiley-Interscience. 2010; 567;Available from: https://doi.org/10.1002/9780470638859.
- Huynh BLC, Duong TH, Tanahashi T, Nguyen KPP. Contribution to the study on chemical constituent of the lichen Parmotrema praesorediosum (Nyl.) Hale, Parmeliaceae. Vietnam Journal of Chemistry. 2010; 48:332-337;.
- Yu X, Guo Q, Su G, Yang A, Hu Z, Qu C, Wan Z, Li R, Tu P, Chai X. Usnic acid derivatives with cytotoxic and antifungal activities from the lichen Usnea longissimi. Journal of Natural Product. 2016; 79: 1373–1380;PMID: 27186821. Available from: https: //doi.org/10.1021/acs.jnatprod.6b00109.