

Evaluation of Microencapsulated Roselle Extract as a Food Colorant in the Red Nata de Coco Dessert Model System

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ABSTRACT

Introduction: The incorporation of natural extracts into food products offers significant health benefits and promotes sustainable production methods. This study evaluated the incorporation of a microencapsulated roselle (*Hibiscus sabdariffa*) extract into nata de coco, a traditional Vietnamese dessert. Anthocyanins, a subgroup of polyphenols, are notable for their vibrant colors and health benefits. By transforming extract solutions into powder forms with maltodextrin and xanthan gum through spray-drying, their color is retained, and their bioavailability is improved, making them effective natural food colorants. **Methods:** The color dyeing ability of the microencapsulated roselle extract was determined on the Natta de coco by direct and indirect dyeing. The investigations on pH, total titratable acidity, anthocyanins, and DPPH were performed on components of the Natta de coco, which is the sugar syrup, and the Natta de coco. Changes in the chemical profiles of the dessert model during storage were also reported in this study. **Discussion:** This study demonstrated that the roselle-enhanced nata de coco exhibited high antioxidant activity and maintained anthocyanin stability. The optimal dyeing method produced samples with a pH ranging from 2.6 to 3.2, a titratable acidity ranging from 0.45% to 0.49%, and a total anthocyanin content ranging from 1.1 to 1.9 mg/L. The degree of antioxidant inhibition was approximately $49.26 \pm 3.0\%$. Stability studies of Roselle Nata de Coco (RDC) over four weeks revealed a consistent pH, titratable acidity, and antioxidant properties with minimal anthocyanin degradation. **Conclusion:** The encapsulated roselle powder enhances the color, nutritional value, and health benefits of nata de coco, demonstrating its potential as a stable natural food colorant. These findings suggest that the encapsulated roselle powder is a stable functional food ingredient with significant potential when combined with Natta de coco. These results highlight the stability and bioactive properties of the encapsulated extract in the food industry, especially for naturally derived and antioxidant-supported products.

Key words: Microencapsulated roselle, anthocyanins, antioxidant, food colorant, nata de coco

1 INTRODUCTION

2 In recent years, plant-based nutritional supplements
3 and plant-based functional foods have gained signifi-
4 cant popularity among individuals seeking to increase
5 their health and well-being^{1,2}. Consequently, inter-
6 est in incorporating pigments from extracts into food
7 matrices to provide safety benefits and nutritional
8 benefits is increasing³⁻⁶. Extracts can enhance the
9 nutritional value and health benefits of food prod-
10 ucts in various forms, including as a food colorant
11 supplement⁴. Anthocyanins, a subgroup of polyphe-
12 nols, are among the most well-known natural sub-
13 strates. Anthocyanins contribute to the vibrant colors
14 observed in various fruits, vegetables, cereals, flow-
15 ers, and other plants^{7,8}. These compounds are found
16 in the flowers, fruits, foliage, roots, tubers, stems,
17 and bulbils of most angiosperms, as well as in some
18 gymnosperms, ferns, and bryophytes. The presence

of anthocyanins can protect against certain cancers⁹,
cardiovascular diseases¹⁰, and other oxidative stress-
related disorders¹¹. Notably, anthocyanins exhibit
significantly greater activity than do common antiox-
idants such as ascorbate¹². Previous studies have
shown that factors such as pH, temperature, and the
presence of light, oxygen, water, or metallic ions (e.g.,
Fe²⁺, Fe³⁺, Ca²⁺, K⁺) significantly influence the sta-
bility of anthocyanins¹³. In addition to their role in
attracting pollinators and assisting in seed dispersal,
dietary anthocyanins have been linked to being nat-
ural food colorants. However, integrating extracts
into food products poses challenges, depending on
the type of food product and requiring precise con-
trol over the extraction and cooking process, which
can degrade the quality and applicability of the plant
extract in food products.

There is considerable interest in incorporating en-
capsulated anthocyanin powders into food products

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38 because of their potential benefits^{14,15}. Encapsu- 92
39 lation powders are produced through encapsulation 93
40 and spray-drying techniques, which transform the 94
41 properties of extract solutions into powder forms¹⁶. 95
42 These techniques protect plant extract components, 96
43 such as polyphenols, anthocyanins, and flavonoids, 97
44 from degradation, retain their vibrant colors, allow 98
45 controlled release for improved bioavailability, mask 99
46 undesirable flavors, and enable their use in a wide 100
47 range of food applications¹⁷. Various studies have
48 explored the use of encapsulated extracts in prod-
49 ucts such as jelly candy¹⁸, yogurt¹⁹, beverages³, and
50 jam²⁰. Encapsulated products not only preserve the
51 core nutritional value of the extract but also offer a
52 solution for quality control, as the powder form al-
53 lows for better measurement and consistency. How-
54 ever, the effectiveness of extraction varies across dif-
55 ferent food models with varying water activities^{21,22}.
56 For high-water activity products such as beverages,
57 juices, or desserts, microencapsulated powders focus
58 on achieving uniform dissolution, maintaining nutri-
59 tional content, and altering the color or flavor of the
60 product.

61 In this study, the encapsulated product from roselle
62 extract was evaluated as a potential food colorant in
63 a high-water-activity food model system. The mi-
64 croencapsulated roselle powder was combined with
65 nata de coco, a traditional Vietnamese dessert, to
66 produce a red nata de coco. Roselle (*Hibiscus sab-*
67 *dariffa*) is known for its high anthocyanin content
68 and is consumed in various forms, including herbal
69 tea and juice^{14,23}. However, the use of roselle ex-
70 tract in solution poses challenges in maintaining sta-
71 ble quality. The microencapsulated powder produced
72 through spray drying offers enhanced stability, allow-
73 ing better control over its properties and increasing
74 the potential of the roselle used in food products.
75 Roselles, which are rich in antioxidants, can be in-
76 corporated into various foods. Recent research has
77 indicated that spray-dried roselle powder can signifi-
78 cantly preserve its antioxidant properties when added
79 to gummies²⁴ and jam products²⁵ and can even pro-
80 long the shelf-life of refrigerated chicken nuggets²⁶.
81 Despite its promising potential, further research is
82 needed to explore its specific application in foods
83 such as nata de coco. By incorporating encapsulated
84 roselle extract into nata de coco, its potential to im-
85 prove bioactive properties, health benefits, and sen-
86 sory attributes could be altered. This study evaluated
87 the dyeing ability and physicochemical properties of
88 red nata de coco, which was created by incorporating
89 microencapsulated hibiscus calyx extract into tradi-
90 tional nata de coco. The preservation of roselle prop-
91 erties such as color stability and bioactive compounds

in high-water-activity products such as nata de coco
was thoroughly analyzed. Combining bioactive com-
pounds from hibiscus with nata de coco is expected to
produce a new food product with enhanced color, nu-
tritional value, and health-promoting properties, po-
tentially replacing commercial food colorants. This
research could lead to the development of promising
functional food products that promote healthier diets
and lifestyles.

MATERIALS & METHODS

Materials

Dry roselle (hibiscus) was purchased from Hichagol
Co., Ltd., Hue, Vietnam. Acetic acid, citric acid,
and sodium hydroxide were purchased from Xilong
Chemical Co. Ltd. Potassium chloride and sodium
acetate anhydrous were purchased from Guangdong
Guanghua Sei-Tech Co. Ltd. Maltodextrin (DE: 10–
12%) was received from Qinhuangdao Lihua Starch
Co. Ltd. DPPH (2,2-diphenyl-1-picrylhydrazyl) and
dimethyl sulfoxide (DMSO) were purchased from
Sigma–Aldrich.

Roselle extraction

The dry roselle flower was ground into a fine powder
with a particle size of approximately 0.5 cm. An acetic
acid 2% extraction solvent was prepared for the ex-
traction process. Roselle powder was immersed in the
solvent at a ratio of 4 g powder to 100 mL extraction
solvent, and the mixture was incubated at 4 °C. After
24 ± 2 h, the extraction mixture was collected, and the
remaining solid residue was subjected to a second ex-
traction under the same conditions. The final roselle
extract was stored at 4 °C for subsequent analysis.

Preparation of spray-dried roselle powder

Before microencapsulation, each encapsulating agent
was dispersed into the roselle extracted solution and
mixed until fully homogenized. The encapsulation
concentration was 10% w/v. The components of the
wall materials used were maltodextrin and xanthan
gum at a 99:1 w/w ratio. The SD-1000 Eyla (EYELA,
Japan) was used to spray the encapsulating mixture.
The homogenized mixture was fed into the spray
dryer, with the inlet temperature set at 160 °C and the
outlet temperature at 80 °C. The solution was fed into
the dryer with a spray air flow rate of 0.7 m³/min and a
pressure drop of 12 × 10 kPa. To maintain the homo-
geneity of the solution, gentle mixing was performed
during the pumping process. After spray drying, the
powders were weighed, stored in airtight plastic bags,
and stored at -20 °C until further evaluation.

141 **Scanning electron microscopy of the spray-**
142 **dried roselle powder**

143 The microstructure of the powders was examined by
144 scanning electron microscopy (SEM). The size and
145 shape of the encapsulated roselle powder particles
146 were determined through image analysis with ImageJ
147 software.

148 **Nata de coco preparation**

149 Dry-pressed raw coconut nata de coco, which was 1.5
150 cm × 1.5 cm in size, was boiled in water containing
151 2% w/v acetic acid and 0.5% w/v iodized salt at 90–
152 95 °C to remove contaminants and residual bacteria.
153 After 40–60 min of boiling, the nata de coco was im-
154 mersed in water for more than 2 h, and the water was
155 changed to remove the acetic acid. After 4–5 cycles of
156 immersion in deionized water, the nata de coco was
157 drained and squeezed to prepare for cooking red nata
158 de coco.

159 **Preparation of Roselle Nata de Coco (RDC)**

160 Microencapsulated roselle powder was used to color
161 nata de coco via two distinct methods: staining via
162 sugar syrup (Method 1 – RDC-1) and staining via
163 direct contact with the nata de coco (Method 2 –
164 RDC-2). The sugar mixture used in both methods
165 contained 26% white sugar (Table 1). Following the
166 staining process, RDC underwent hot packaging and
167 was kept in a refrigerator until further characteriza-
168 tion. The processing scheme of the two staining meth-
169 ods is presented in Figure 1. White nata de coco
170 (nonstaining sample) and commercial colorant (ama-
171 ranth) samples were prepared under the same condi-
172 tions for comparison.

173 **Physicochemical profile of RDC**

174 **pH determination and titratable acidity**

175 The titratable acidity was determined following the in-
176 structions of Tyl and Sadler, 2009, in the Food Analy-
177 sis book²⁷. Five milliliters of RDC was added to 45
178 mL of distilled water. The pH of the solution was
179 measured. The samples were titrated by adding 0.1
180 N NaOH to the mixture in the presence of phenolph-
181 thalein until the pH reached 7.0. The titratable acidity
182 of RDC was calculated via the following equation:

183 % titratable acidity (wt/wt) = $N \times V \times \text{Eq.wt.}$
184 $W \times 1000 \times 100$

185 where: N = normality of the titrant, usually NaOH
186 (mEq/mL)

187 V = volume of titrant (mL)

188 Eq.wt. = equivalent weight of predominant acid
189 (mg/mEq)

W = mass of sample (g) 190
1000 = factor relating mg to grams (mg/g) (1/10 = 191
100/1000) 192

193 **Total anthocyanin content**

194 The anthocyanin content was determined according
195 to the procedure of Lee et al. (2005), with slight mod-
196 ifications²⁸. Two different buffers were prepared, in-
197 cluding a pH 1.0 buffer (potassium chloride, 0.025 M)
198 and a pH 4.5 buffer (sodium acetate, 0.4 M). The sam-
199 ple was mixed with each buffer at a ratio of 1:4. The
200 absorbance of each of these solutions was measured
201 at 520 nm and 700 nm via a microplate reader and a
202 UV spectrophotometer. The absorption difference of
203 the attenuated sample was computed according to the
204 following equation:

205 $A = (A_{520} - A_{700})_{\text{pH 1.0}} - (A_{520} - A_{700})_{\text{pH 4.5}}$

206 The total anthocyanin concentration was expressed as
207 cyanidin-3-glucoside equivalents (% w/w) via the fol-
208 lowing formula:

209 $\text{Anthocyanin [mg/L]} = A \times \text{MW} \times \text{DF} \times 1000 \epsilon \times 1$

210 where A = absorbance

211 MW = molecular weight (449.2 g/mol)

212 DF = dilution factor

213 ϵ = extinction molar coefficient (26,900 L/cm.mol)

214 **Antioxidant assay – DPPH assay**

215 DPPH was prepared at 0.1 mM in a DMSO solu-
216 tion. The RDC in each cooking method, nata de coco
217 without spray-dried roselle extract as a control, spray-
218 dried roselle extract in distilled water, and DMSO as
219 a blank were sampled. Different concentrations of
220 samples were diluted in DMSO. A total of 180 μL of
221 0.1 mM DPPH solution was incubated with a 20 μL
222 sample for 30 minutes in the dark. A series of ascor-
223 bic acid (VitC) dilutions in DMSO at concentrations
224 ranging from 0–200 μg/mL were also prepared and
225 measured at the same time. The decrease in the ab-
226 sorbance of DPPH was measured at 517 nm via a mi-
227 croplate reader. The proportion of the scavenging ef-
228 fect of DPPH was calculated via the following equa-
229 tion:

230 $\text{DPPH scavenging effect (\%)} = [(A_0 - A_1) / A_0] \times 100$

231 where A0 = the absorbance of the control.

232 A1 = the absorbance of the sample

233 **Storage stability**

234 RDC was prepared and packaged in 150 mL plastic
235 sealed bags. For storage ability analysis, RDC sam-
236 ples were stored for 4 weeks in a refrigerator (4 °C)
237 without light. For quantitative analysis, the pH, titrat-
238 able acidity, total anthocyanin content, and antioxi-
239 dant activity of the samples were determined via the

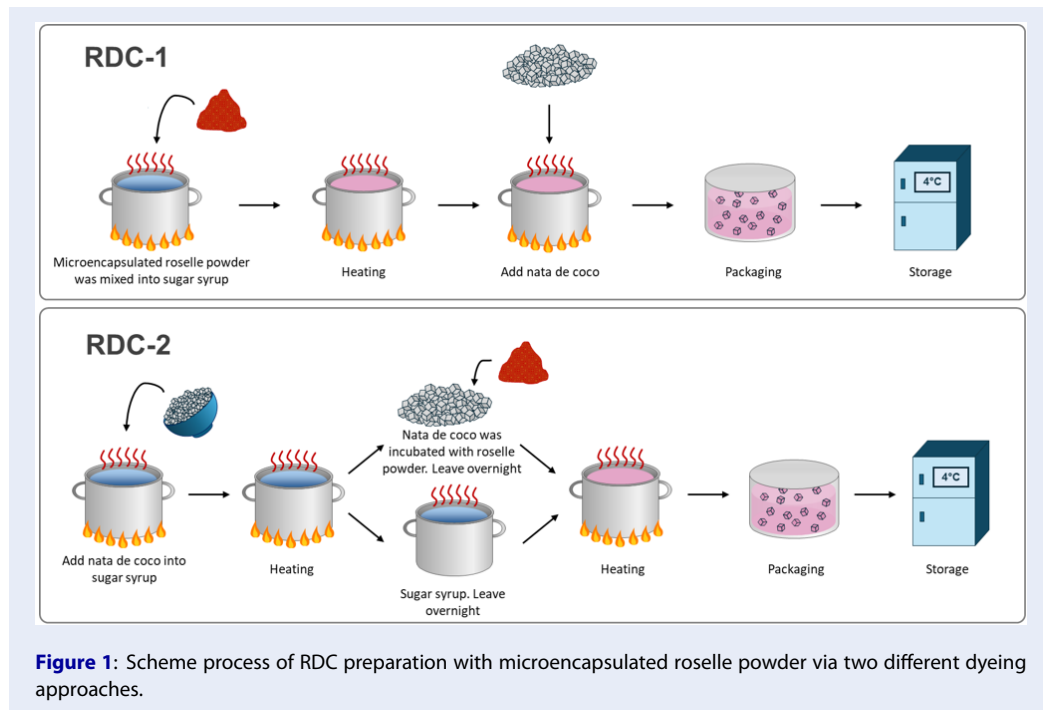


Figure 1: Scheme process of RDC preparation with microencapsulated roselle powder via two different dyeing approaches.

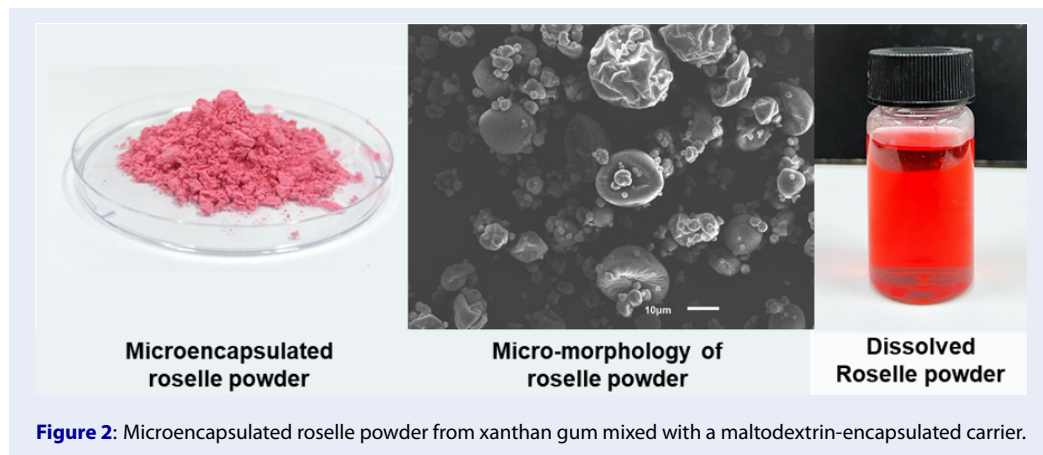


Figure 2: Microencapsulated roselle powder from xanthan gum mixed with a maltodextrin-encapsulated carrier.

Table 1: Ingredients of RDC.

INGREDIENTS	RDC	White nata de coco	Amaranth nata de coco
Sugar concentration (w/w):	26%	26%	26%
Sugar (°Brix):	21 °Brix	21 °Brix	21 °Brix
Nata de coco: syrup ratio:	60:40	60:40	60:40
Roselle concentration (w/w):	3%	-	-
Commercial colorant (w/w)	-	-	0.5

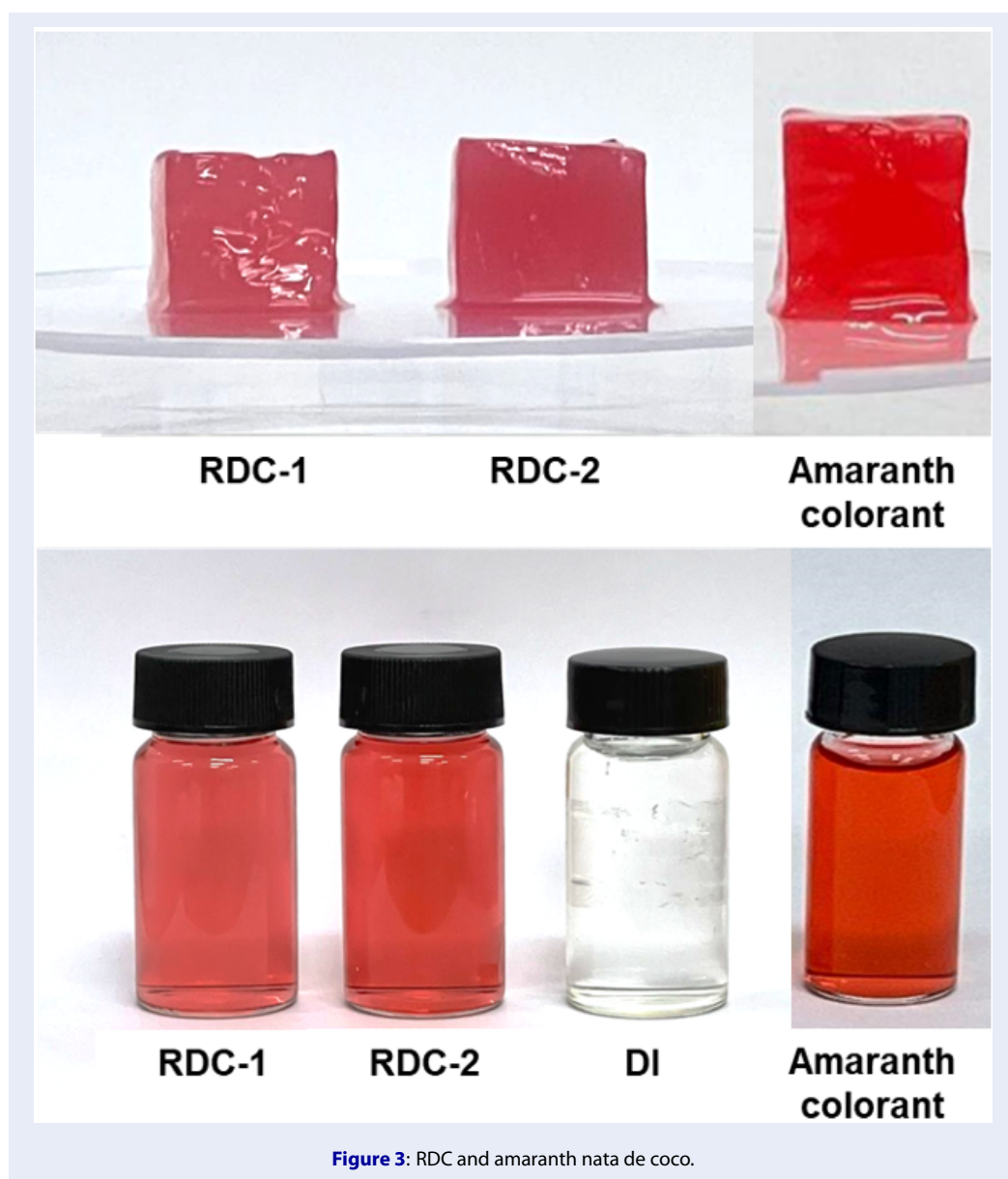


Figure 3: RDC and amaranth nata de coco.

240 DPPH method to evaluate the changes in the degra-
 241 dation of the sample characteristics each week.

242 **Statistical analysis**

243 All the results are presented as the means \pm standard
 244 deviations. Statistical analysis was conducted via one-
 245 way ANOVA for all tests, followed by Tukey's post hoc
 246 analysis to determine significant differences between
 247 groups. A p value of less than 0.05 was considered
 248 statistically significant.

249 **RESULTS**

Microencapsulated roselle powder

250 The spray-dried hibiscus extract powder exhibited
 251 light pink coloration, combining the characteristic red
 252 hue from the hibiscus with the white tone of the en-
 253 capsulated carrier. The powder was a fine, particu-
 254 late powder comprising small, granular particles. Ini-
 255 tially, upon testing the solubility of the powder, the
 256 microencapsulated powder exhibited 70% solubility.
 257 The anthocyanin content and antioxidant properties
 258 determined via DPPH were 25.07 mg/mL. Accord-
 259 ing to the SEM results, the microencapsulated roselle
 260 powder had a circular morphology with rough and
 261 wrinkled surfaces. The microparticles were approxi-
 262

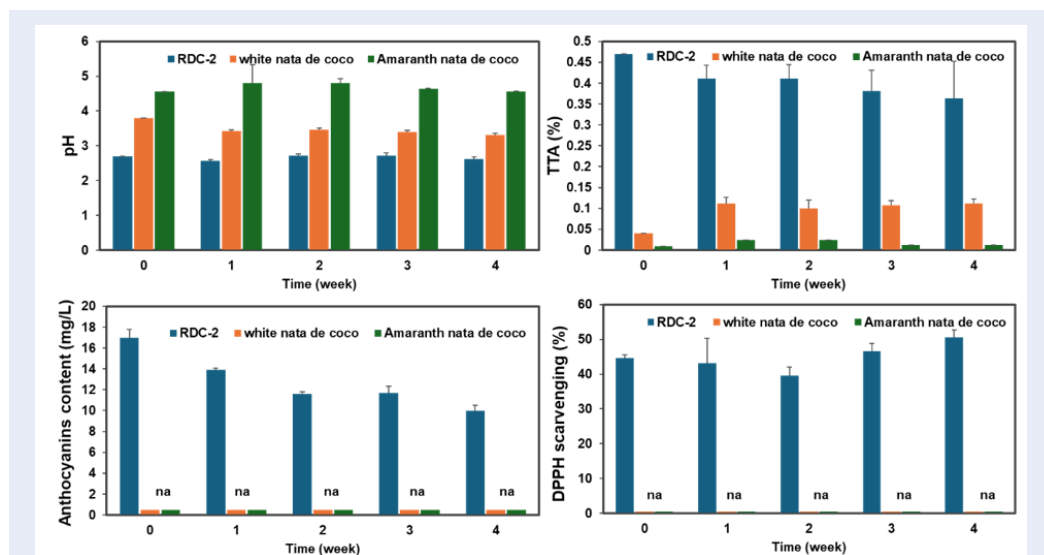


Figure 4: Physicochemical properties of RDC, white nata de coco, and amaranth nata de coco during four weeks of storage in a refrigerator.

Table 2: Physicochemical profile of nata de coco produced under different treatments.

	pH	TTA (%)	Anthocyanin content (mg/L)	DPPH scavenging (%)	µgVitC/mL
Sugar syrup					
RDC-1	2.6 ± 0.0	0.49 ± 0.00	1.17 ± 0.08 ^c	41.00 ± 2.37 ^b	115.67 ± 6.78 ^b
RDC-2	2.7 ± 0.0	0.47 ± 0.00	1.70 ± 0.11 ^b	44.57 ± 0.97 ^b	126.01 ± 2.78 ^b
White nata de coco	3.8 ± 0.0	0.04 ± 0.00	-	-	-
Roselle powder in DI	2.9 ± 0.1	0.47 ± 0.00	2.26 ± 0.00 ^a	57.45 ± 1.80 ^a	162.86 ± 5.16 ^a
Amaranth nata de coco	4.56 ± 0.0	0.01 ± 0.00	-	-	-
Nata de coco					
RDC-1	3.2 ± 0.5	0.45 ± 0.02	1.15 ± 0.09 ^c	42.85 ± 4.8 ^c	115.49 ± 9.62 ^c
RDC-2	2.7 ± 0.0	0.47 ± 0.01	1.92 ± 0.07 ^b	49.26 ± 3.0b ^c	143.27 ± 4.88 ^b
White nata de coco	3.9 ± 0.0	-	-	-	-
Roselle powder in DI	2.9 ± 0.1	0.47 ± 0.00	2.26 ± 0.00 ^a	57.45 ± 1.80 ^a	162.86 ± 5.16 ^a
Amaranth nata de coco	4.3 ± 0.1	0.01 ± 0.00	-	-	-

*Different letters represent significantly different data

mately 15 μm in size. This morphology indicated successful microencapsulation using maltodextrin, xanthan gum, and hibiscus extract. Figure 2 presents the characteristics of the roselle encapsulated powder.

Red RDC

To produce red-colored nata de coco, two staining approaches were tested to evaluate the effectiveness of microencapsulated roselle powder as a food coloring agent. The powder was stained indirectly by mixing with syrup via one method and directly with nata de coco via the other method. Both techniques resulted in noticeable color heterogeneity in the nata de coco. Method 2, where the powder was added to the nata de coco, produced a slightly darker hue than did Method 1, where the powder was added to the syrup. This color difference is attributed to the timing of the addition of the spray-dried roselle extract powder during the cooking process. The morphology of the red-colored nata de coco (RDC) produced by both methods is shown in Figure 3.

The physicochemical properties of the syrup and nata de coco are presented in Table 2. To assess the staining efficiency of the encapsulated roselle powder in RDC, the physico-chemical profiles of RDC, microencapsulated roselle powder in deionized water (DI), and a blank sample without a roselle were analyzed. Table 2 presents the differences in properties under various treatments. In the sugar mixture, the pH of the RDC produced via both methods was approximately 2.6. However, in nata de coco, the pH of Method 2 (2.7) was lower than that of Method 1 (3.2). This may be due to the complexity of the cellulose matrix of the nata de coco, which does not easily enable the extract to be absorbed. This finding indicated that directly introducing roselle powder into drained nata de coco enhances the absorption of the extract. Conversely, the titratable acidity between Method 1 and Method 2 (0.45–0.49%) showed minimal differences.

The anthocyanin content and antioxidant activity are crucial parameters for evaluating the functionality of roselle powder in nata de coco. Both metrics were higher in the samples from Method 2 (RDC-2) than in those from Method 1 (RDC-1). Specifically, RDC-2 had anthocyanin concentrations of 1.7 mg/L in syrup and 1.93 mg/L in nata de coco, whereas the RDC-1 samples contained approximately 1.15 to 1.17 mg/L. This trend was also observed for the antioxidant activity, suggesting that adding roselle to drained nata de coco and incubating overnight enhances the incorporation and diffusion into both nata de coco and sugar syrup. Compared with the sample of roselle powder

dissolved in DI water, the RDC samples had minimal effects on the cooking process. Moreover, no anthocyanins or antioxidant activities were detected in samples colored with a commercial amaranth colorant, further highlighting the functional benefits of encapsulated roselle powder.

In the stability investigation, the physicochemical properties of RDC stored at 6 ± 2 °C for four weeks were evaluated to monitor potential changes over time. The pH of the products did not change over four weeks, with the pH ranging from 2.5 to 4.0 under both storage conditions. Similarly, the titratable acidity (TTA) exhibited minimal variation. However, as shown in Figure 4, the anthocyanin content in each sample decreased considerably by approximately 10–20% compared with that in the previous week. Despite the decrease in anthocyanin content, Figure 4 shows that the antioxidant properties of each sample remained approximately the same. These findings suggest that antioxidant activity is not solely dependent on anthocyanin levels.

DISCUSSION

Plant pigments have long been used as natural food colorants in the culinary industry. When applied to nata de coco, microencapsulated roselle powder can increase the demand for roselle, potentially increasing production and benefiting the economies of the roselle- and nata de coco-producing regions. Producers can achieve greater profits by incorporating spray-dried roselle into their products. The beneficial use of roselle in nata de coco reduces the reliance on synthetic food colorants. Consumers benefit from the added nutritional value and reduced risk of allergic reactions or problems caused by commercial colorants. This natural alternative to synthetic colorants also provides antioxidants and anti-inflammatory compounds, enhancing the health benefits of the product. While it may not serve as a treatment for specific conditions, it offers a healthful alternative to synthetic food colorants, positively impacting consumer health.

Encapsulation involves surrounding a central material with a wall material to prevent chemical and environmental interactions²⁹. This method addresses the instability of substances by inhibiting volatilization and protecting against adverse conditions³⁰. In this study, roselle powder was encapsulated with maltodextrin and xanthan gum. The concentration of xanthan gum is crucial for maintaining the antioxidant and color properties of roselle extracts. The characteristics of the encapsulated powder are shown in

365 Figure 2. The combination of plant extracts with mal-
366 to-dextrin and xanthan gum effectively provided color
367 and antioxidant properties. Thus, microencapsulated
368 granules under these conditions are a viable option for
369 developing functional foods.

370 In combining and developing nata de coco products
371 by using roselle encapsulated powder, understanding
372 the characteristics of nata de coco is essential. Cel-
373 lulose, a natural polymer composed of carbohydrate
374 polysaccharides, is obtained via fermentation and ad-
375 heres to the cellulose fibers produced by *Acetobacter*
376 *xylinum*. Physical characteristics can affect food qual-
377 ity and consumer acceptability. Nata de coco has high
378 aesthetic value because of its pure white hue, dense
379 consistency, and fresh scent³¹. To produce bacte-
380 rial cellulose for nata de coco, the product is thor-
381 oughly rinsed to remove the sour taste and then boiled
382 to eliminate the acidic flavor and eliminate any mi-
383 croorganisms. After purification, the bacterial cellu-
384 lose is cooked with sugar and other ingredients to cre-
385 ate a dessert known as nata de coco. In this study,
386 microencapsulated roselle powder was used as a col-
387 orant to introduce a safe, aesthetically appealing, and
388 antioxidant-rich nata de coco. Given that water activ-
389 ity is crucial in regulating pigment degradation dur-
390 ing storage, this study aims to understand the protec-
391 tive effect of encapsulation on roselle extract in a high-
392 water-activity food model such as nata de coco.

393 The ability of the microencapsulated powder as a col-
394 orant for nata de coco was assessed via two different
395 methods. In Method 1, roselle powder was added
396 during the initial cooking phase when the nata de
397 coco remained porous and could absorb liquid. The
398 structural matrix of the gel likely absorbed some phy-
399 tochemicals, resulting in less color in the residual
400 syrup. In contrast, in Method 2, roselle was added
401 after cooking the nata de coco, leaving more extract
402 particles suspended in the sugar syrup rather than ab-
403 sorbed into the nata de coco fibers. The dispersed
404 pigments in method 2 imparted a more intense red
405 coloration to the solution. The results indicated that
406 encapsulated roselle powders exhibited their primary
407 ability to stain the subject directly or indirectly after
408 being dissolved in a solution.

409 Titratable acidity and pH offer complementary in-
410 sights into food quality, with titratable acidity more
411 accurately reflecting the impact of organic acids on
412 flavor. Food acids only partially ionize, affecting some
413 properties on the basis of the ionized fraction and
414 others on the total acid content. pH determination
415 is crucial for developing high-quality products with
416 optimal phytochemical characteristics³². According
417 to Paramee Chumsri et al., the optimal pH of dried

roselle calyxes in beverages ranges from 2.8 to 2.9³³.
Similarly, the pH of RDC in this study was approx-
imately 2.6–3.2, making the product sour due to the
acidity of the encapsulated roselle powder. The acid-
ity level of spray-dried RDC is approximately 0.4%.
The antioxidant properties were approximately 50%
in both samples. These findings indicate that spray-
dried roselle powder is appropriate for use in food and
beverages. Using spray-dried roselle in nata de coco
can reduce the need for synthetic food colorants, of-
fering a natural and healthy alternative.

The total anthocyanin content serves as an indicator
of both the color and potential health benefits pro-
vided by the anthocyanins in roselle extract. The
results revealed that Method 2 had a significantly
greater anthocyanin content (1.7–1.93 mg/L) than
Method 1 did (1.15–1.17 mg/L). In Method 1 (RDC-
1), adding the spray-dried extract while cooking with
sugar syrup allowed more roselle to remain suspended
in the residual syrup rather than being absorbed into
the nata de coco. Conversely, in Method 2 (RDC-
2), the roselle powder was added during the cooking
of nata de coco, which has significant water-holding
capacity, enabling a portion of the anthocyanins to
be absorbed into the cellulose matrix of the nata de
coco. This is visually evident, as the color of the
sugar syrup in Method 2 is slightly less red than that
in Method 1, but the nata de coco in Method 2 is
redder. The two methods assess a sample's antioxi-
dant activities to determine the anthocyanin antioxi-
dant outcome following nata de coco production.
This disparity likely arises from differences in pro-
cessing conditions. In RDC-1, the nata de coco ex-
posed the extract to potentially longer temperature
effects, decreasing anthocyanin stability and activ-
ity. However, in RDC-2, the addition of roselle ex-
tract after cooling prevented exposure to sudden tem-
perature changes during prolonged cooking. As de-
scribed in the section on total anthocyanin content,
the heating procedure affects the stability of antho-
cyanins^{22,34,35}, leading to a decrease in their antioxi-
dant activity in nata de coco. Furthermore, antho-
cyanins degrade in response to various factors, in-
cluding pH, temperature, intermolecular pigmenta-
tion, ascorbic acid, oxygen, etc.³⁶. The cleavage of
the intramolecular bonds of anthocyanins, resulting
in colorless, smaller molecules, is more active in the
extracts than in the cells because of the high tem-
perature. For example, coumarin glycoside has been
identified as a common byproduct of the thermal
degradation of cyanidin 3,5-diclycosides over time³⁷.
Specifically, at 37 °C, delphinidin 3-sambubioside
and cyanidin 3-sambubioside in anthocyanins lead

471 to the formation of colorless molecules such as gallic
472 acid and protocatechuic acid³⁸. This explains why the
473 anthocyanin content in RDC-1 was lower than that in
474 RDC-2, and the color was also brighter red in RDC-1
475 than in RDC-2. The effects of temperature on antho-
476 cyanin properties were also observed when the antho-
477 cyanin contents produced in treatments 1 and 2 were
478 compared with those of the same amount of roselle
479 powder mixed with DI. After processing, the medic-
480 inal properties were reduced by approximately 30%
481 compared with those of the original product.

482 In the stability study, the physicochemical properties
483 of the RDC samples remained stable over four weeks,
484 except for the anthocyanin content. The stability of
485 anthocyanins in roselle powder is influenced by vari-
486 ous factors, such as heat and pH, particularly in low-
487 acid environments where anthocyanins degrade more
488 rapidly. However, the study revealed that the antho-
489 cyanins in the roselle powder presented high color
490 stability and heat resistance under acidic conditions.
491 These findings suggest that the quality and appearance
492 of roselle powder can be preserved by adjusting the
493 pH and temperature during processing. Furthermore,
494 the roselle extract used in this study was rich in antho-
495 cyanins and exhibited strong antioxidant properties.
496 Despite a decrease in anthocyanin levels over time, the
497 antioxidant activity remained stable, further support-
498 ing the potential of roselle powder as a food ingredi-
499 ent with health benefits. Some studies have indicated
500 that roselle anthocyanins are relatively stable at high
501 temperatures, particularly when the heating duration
502 is short (30–45 min)^{39,40}. The preservation of roselle
503 properties in a high-water-activity food model such
504 as nata de coco demonstrates the potential of roselle-
505 encapsulated powder as a natural colorant. Therefore,
506 further research is warranted to determine the opti-
507 mal conditions for encapsulating roselle anthocyanins
508 during the heating process. Importantly, this study
509 assessed the stability of anthocyanins in roselle pow-
510 der under specific conditions, and the results may not
511 be applicable to other anthocyanin-containing prod-
512 ucts or processing methods. Additional research is
513 needed to explore the stability of anthocyanins in var-
514 ious food and beverage products and their potential
515 applications.

516 CONCLUSIONS

517 In conclusion, this study successfully developed a nata
518 de coco product using encapsulated roselle powder
519 as a natural food colorant. The results demonstrated
520 high antioxidant activity and maintained the stabil-
521 ity of the anthocyanins in the RDC product. The opti-
522 mal method involved incubating roselle in drained

nata de coco, producing samples with pH values rang- 523
ing from 2.6–3.2, titratable acidities between 0.45% 524
and 0.49%, and total anthocyanin contents varying 525
from 1.1–1.9 mg/L. The degree of antioxidant inhibi- 526
tion ranged from 40–50%. Stability studies confirmed 527
that the pH, titratable acidity, and antioxidant prop- 528
erties of the samples remained consistent over four 529
weeks, despite some anthocyanin degradation. This 530
research explored the potential of a natural food col- 531
orant to create an innovative product, offering valu- 532
able insights for future research and development. By 533
providing a low-cost and effective natural alterna- 534
tive to synthetic colorants, this approach may posi- 535
tively impact consumer health. These findings sug- 536
gest that the use of microencapsulated RDC powder 537
can lead to the production of stable functional foods 538
with significant potential. Future research should pri- 539
oritize the assessment of RDC stability under various 540
storage conditions and conduct broader sensory eval- 541
uations of consumer preferences. Although roselle 542
powder provides antioxidants and anti-inflammatory 543
compounds, potentially improving health benefits for 544
traditional nata de coco, further research is needed 545
to fully understand the health benefits of spray-dried 546
roselle. Once the functional food's suitability and 547
health benefits are confirmed, further in vivo and in 548
vitro studies will be conducted to evaluate its efficacy. 549
These investigations provide crucial insights into the 550
potential culinary and health applications of RDC. 551

552 LIST OF ABBREVIATIONS USED

DPPH: 2,2-diphenyl-1-picrylhydrazyl 553
DMSO: Dimethyl sulfoxide 554
RDC: Roselle Nata de Coco 555
TTA: titratable acidity 556

557 COMPETING INTERESTS

The authors declare that they have no competing inter- 558
ests. 559

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567 AUTHORS' CONTRIBUTION

Xxx 568

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