

# Phenolic compounds from the bark of *Aglaia lawii* and their cytotoxic activity against HepG2

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## ABSTRACT

**Introduction:** *Aglaia lawii* is a large tree belonging to the Meliaceae family. In traditional medicine, stem bark is used as a vermicide, whereas leaves are used for the treatment of headache. Studies on the chemical constituents of this species in Vietnam and its biological activity, especially its ability to treat liver cancer, are still limited. This work describes the isolation and structural elucidation of six phenolic compounds from an ethyl acetate extract of the bark of *A. lawii* as well as the evaluation of their cytotoxic activity against HepG2. **Methods:** Extracts were prepared by extracting dried ground bark via Soxhlet extraction. Isolation was performed via column chromatography over silica gel, RP-18, and via gel permeation chromatography over Sephadex LH-20. Chemical structures were elucidated via spectroscopic methods (1D- and 2D-NMR, HR-ESI-MS and IR), and the spectral data were compared with those in the literature. Cytotoxic activity against HepG2 human liver cancer cells was evaluated in vitro via the MTT assay. **Results:** Six phenolic compounds, palmarumycin JC2 (**1**), 5-hydroxy-4',7-dimethoxyflavone (**2**), coniferaldehyde (**3**), 3-hydroxy-1-(4'-hydroxy-3'-methoxyphenyl)propan-1-one (**4**), *p*-hydroxybenzaldehyde (**5**), and vanillin (**6**), were isolated from an ethyl acetate extract of the bark of *A. lawii* collected in Dong Nai Province. Compounds **1-3** were evaluated for their cytotoxicity against HepG2 cells, and the results showed that the compounds exhibited weak effects or no activity. **Conclusion:** All the isolated compounds have been reported from this species for the first time. This is also the first report on the cytotoxicity of **1-2** against HepG2 cells, although the compounds displayed weak effects.

**Key words:** *Aglaia lawii*, phenolic compounds, cytotoxicity, HepG2

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## 1 INTRODUCTION

**2** *Aglaia* is the largest genus of the Meliaceae family and  
**3** consists of approximately 250 species<sup>1</sup>. *Aglaia lawii*  
**4** (Wight) Fald. ex Raman<sup>2</sup> or *A. lawii* (Wight) C.J.  
**5** Saldanha (synonyms: *Aglaia andamanica*, *Amoora*  
**6** *tsangii*)<sup>3</sup> is found mainly in India, Bhutan, China,  
**7** and Southeast Asia, including Vietnam, Myanmar,  
**8** Thailand, Indonesia, Malaysia, the Philippines, and  
**9** Laos<sup>1</sup>. In folk medicine, the leaves are used for  
**10** the treatment of headache<sup>2</sup>, and the stem bark is  
**11** used as a vermicide<sup>4</sup>. Previous phytochemical in-  
**12** vestigations of this species have revealed the sig-  
**13** nificant existence of limonoids<sup>4-6</sup>, triterpenoids<sup>7,8</sup>,  
**14** sesquiterpenoids<sup>5,9</sup>, and steroids<sup>10</sup>, which exhibit di-  
**15** verse pharmacological activities, such as cytotoxic<sup>7</sup>,  
**16** anti-inflammatory<sup>9-11</sup>, antiallergic<sup>11</sup>, and anti-HIV-  
**17** 1<sup>12</sup> properties. In this work, we report the isolation  
**18** and structural characterization of six phenolic com-  
**19** pounds (**1-6**) from the ethyl acetate of the bark of  
**20** *Aglaia lawii*. Furthermore, the in vitro cytotoxic activ-  
**21** ity of the isolated compounds against HepG2 human  
**22** liver cancer cells was evaluated via the MTT assay to

identify active compounds with therapeutic effects on  
liver cancer.

## MATERIALS AND METHODS

### General experimental procedures

Optical rotation was measured via a P8000 polarime-  
ter manufactured by A. Krüss Optronic, whereas HR-  
ESI-MS data were recorded via an Agilent 6500 series  
Q-TOF mass spectrometer. IR spectra were recorded  
with KBr using a JASCO FT/IR-6600 spectrometer.  
NMR spectra were obtained on a Bruker AV 500 (500  
MHz for <sup>1</sup>H and 125 MHz for <sup>13</sup>C) with CDCl<sub>3</sub>,  
acetone-*d*<sub>6</sub> or methanol-*d*<sub>4</sub> as the solvent and cali-  
brated on the basis of the chemical shifts of the corre-  
sponding deuterated solvents<sup>13</sup>. Column chromatog-  
raphy (CC) was run on silica gel (Merck, 40–63 mm)-  
or RP-18 (Merck, 40–63 mm)-bonded phases. For gel  
permeation chromatography (GPC), Sephadex LH-20  
(GE Healthcare) was used. TLC was carried out on  
TLC silica gel (Merck, 250 mm)- or RP-18 (Merck,  
200 mm)-precoated aluminum plates. The TLC plates  
were visualized via UV light, sprayed with ethanolic

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44 ferric chloride or vanillin-H<sub>2</sub>SO<sub>4</sub> in EtOH, and then  
 45 heated at approximately 120 °C for several minutes.  
 46 Fetal bovine serum (FBS), minimal essential medium  
 47 with Eagle salt (MEME), and ellipticine were pur-  
 48 chased from Sigma. HepG2 cells (HB-8065<sup>TM</sup>) were  
 49 obtained from the American Type Culture Collection.  
 50 Cytotoxic activity was determined in Costar 96-well  
 51 plates.

## 52 Plant material

53 The bark of *A. lawii* was collected at the Center for  
 54 Experimental Forest Research in Eastern South Viet-  
 55 nam (formerly known as Trang Bom Plant Collection  
 56 Garden), Dong Nai Province, in November 2019. Ac-  
 57 cording to the document kept at the Center, the plant  
 58 has been cultivated at Lot H with Code Number 83,  
 59 Vietnamese name: Gội bốn cánh.

## 60 Extraction and isolation

61 The dried, ground bark (10 kg) was extracted with  
 62 EtOAc and MeOH, respectively, via a Soxhlet extrac-  
 63 tor. Removing the solvents via a rotary evaporator  
 64 produced an EtOAc extract (110 g) and a MeOH ex-  
 65 tract (138 g). The CC of the EtOAc extract with sil-  
 66 ica gel (*n*-hexane-EtOAc 0–100%) furnished 10 frac-  
 67 tions (F1–10). Fraction F6 (9.5 g) was separated via  
 68 CC on silica gel (*n*-hexane-EtOAc 0–50%) to afford  
 69 8 fractions (F6.1–8). Further separation of fraction  
 70 F6.2 (1.22 g) via CC on silica gel (*n*-hexane-CHCl<sub>3</sub>  
 71 30–100%) afforded eight fractions (F6.2.1–8). frac-  
 72 tion F6.2.5 (30.5 mg) was purified via CC on RP-18  
 73 (70–100% MeOH in H<sub>2</sub>O) to give **2** (5.4 mg). frac-  
 74 tion F6.3 (835 mg) was separated via repeated CC on  
 75 silica gel (*n*-hexane-EtOAc 0–70%) to obtain **1** (24.5  
 76 mg). F6.4 (600 mg) was subjected to Sephadex LH-20  
 77 (CHCl<sub>3</sub>-MeOH 1:1) and then purified via CC on sil-  
 78 ica gel (*n*-hexane-EtOAc 0–40%) to produce **3** (12.6  
 79 mg) and **4** (3.5 mg). F6.6 (1.64 g) was subjected to  
 80 CC on silica gel (*n*-hexane-acetone 0–40%) to furnish  
 81 five fractions (F6.6.1–5). The purification of fraction  
 82 F6.6.2 (83.0 mg) via Sephadex LH-20 (CHCl<sub>3</sub>-MeOH  
 83 1:1) yielded **5** (7.2 mg). The CC of fraction F6.6.4  
 84 (55.0 mg) on silica gel (*n*-hexane-EtOAc, 0–40%) led  
 85 to the isolation of **6** (4.4 mg).

## 86 Cytotoxicity assay

87 The MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-  
 88 diphenyltetrazolium) assay was carried out via the  
 89 method described by Mosmann<sup>14</sup>, with ellipticine  
 90 used as the positive control (n = 3). The cells were  
 91 cultured in MEME supplemented with 10% FBS at  
 92 37°C in a humidified atmosphere with 5% CO<sub>2</sub>.

93 Ellipticine was dissolved in DMSO at a concentration  
 94 of 0.01 mM. Briefly, HepG2 cells were separated with  
 95 trypsin, seeded in each well at 3 × 10<sup>4</sup> cells/mL,  
 96 treated with the test compounds at concentrations of  
 97 256, 64, 16, 4 and 1 μg/ml for 72 hours under stan-  
 98 dard conditions, and then stained with 10 μl of MTT  
 99 (5 mg/ml) for 4 hours. The formazan crystals were  
 100 dissolved in 100 μl of DMSO after the environmental  
 101 solutions were removed. Optical density (OD) values  
 102 were measured at a wavelength of 540 nm via a  
 103 96-well microtiter plate reader (Synergy HT, Biotek  
 104 Instruments). The percentage of growth inhibition  
 105 (I%) was calculated according to the formula:

$$I\% = \frac{|OD_c - OD_t|}{OD_c} \times 100$$

106 OD<sub>t</sub>: Optical density values of the tested sample  
 107 OD<sub>c</sub>: Optical density values of the control sample.  
 108 The IC<sub>50</sub> values were calculated via nonlinear regres-  
 109 sion via RawData software (Gen5 2.07.17).

## RESULTS

110  
 111 From the ethyl acetate extract of the bark of *A. lawii*,  
 112 six compounds (**1–6**) were isolated via chromato-  
 113 graphic methods. Their structures are shown in Fig-  
 114 ure 1.

115 **Palmaramycin JC2 (1)**: yellow needles.  $[\alpha]_D^{25} +142.4$   
 116 (*c* 7 mg in 1 ml CHCl<sub>3</sub>); IR (KBr)  $\nu_{max}$ : 3452,  
 117 3059, 2924, 1643, 1612, 1554, 1269, 814, 756 cm<sup>-1</sup>.  
 118 HR-ESI-MS: *m/z* 333.0758 [M-H]<sup>-</sup> (calculated for  
 119 C<sub>20</sub>H<sub>13</sub>O<sub>5</sub> 333.0768). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 500 MHz):  
 120  $\delta_H$  13.84 (1H, s, 9-OH), 7.55 (3H, m, H-7, H-1', H-  
 121 9'), 7.49 (1H, t, 8.0 Hz, H-2'), 7.44 (1H, t, 7.9 Hz, H-  
 122 8'), 7.33 (1H, dd, 7.6, 1.0 Hz, H-6), 7.08 (1H, d, 8.3  
 123 Hz, H-8), 7.08 (1H, d, 8.3 Hz, H-3'), 6.92 (1H, d, 7.5  
 124 Hz, H-7'), 4.60 (1H, t, 3.7 Hz, H-3), 3.24 (1H, dd,  
 125 17.8, 3.4 Hz, H-2a), 2.95 (1H, dd, 17.8, 4.0 Hz, H-2b);  
 126 <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 125 MHz):  $\delta_C$  201.1 (C-1), 162.3  
 127 (C-9), 147.3 (C-6'), 146.5 (C-4'), 138.1 (C-5), 134.3  
 128 (C-10'), 137.2 (C-7), 127.9 (C-2'), 127.8 (C-8'), 121.3  
 129 (C-9'), 121.6 (C-1'), 120.0 (C-8), 118.1 (C-6), 115.5  
 130 (C-10), 113.3 (C-5'), 109.7 (C-3'), 109.0 (C-7'), 98.9  
 131 (C-4), 67.4 (C-3), 41.4 (C-2) (<sup>1</sup>H- and <sup>13</sup>C-NMR as-  
 132 signments were performed using HSQC, HMBC and  
 133 COSY techniques).

134 **5-Hydroxy-4',7-dimethoxyflavone (2)**: yellow nee-  
 135 dles. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta_H$  12.80 (1H, s,  
 136 5-OH), 7.84 (2H, d, 8.9 Hz, H-2'/H-6'), 7.01 (2H, d,  
 137 8.9 Hz, H-3'/H-5'), 6.57 (1H, s, H-3), 6.48 (1H, d, 2.2  
 138 Hz, H-8), 6.36 (1H, d, 2.2 Hz, H-6), 3.89 (3H, s, 7-  
 139 OCH<sub>3</sub>), 3.88 (3H, s, 4'-OCH<sub>3</sub>); <sup>13</sup>C-NMR (CDCl<sub>3</sub>,  
 140 125 MHz):  $\delta_C$  182.6 (C-4), 165.6 (C-7), 164.2 (C-  
 141 2), 162.8 (C-4'), 162.4 (C-5), 157.9 (C-9), 123.8 (C-  
 142 1'), 128.2 (C-2'/C-6'), 114.7 (C-3'/C-5'), 105.8 (C-10),

143 104.5 (C-3), 98.2 (C-6), 92.8 (C-8), 55.9 (4'-OCH<sub>3</sub>),  
144 55.7 (7-OCH<sub>3</sub>).

145 **Coniferaldehyde (3)**: yellowish needles. <sup>1</sup>H-NMR  
146 (CDCl<sub>3</sub>, 500 MHz):  $\delta_H$  9.65 (1H, d, 7.7 Hz, H-9),  
147 7.40 (1H, d, 15.8 Hz, H-7), 7.12 (1H, dd, 8.2, 1.9 Hz,  
148 H-6), 7.07 (1H, d, 1.9 Hz, H-2), 6.96 (1H, d, 8.2 Hz,  
149 H-5), 6.60 (1H, dd, 15.8, 7.7 Hz, H-8), 3.95 (3H, s,  
150 3-OCH<sub>3</sub>); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 125 MHz):  $\delta_C$  193.9  
151 (C-9), 153.4 (C-7), 149.3 (C-3), 147.4 (C-4), 127.1 (C-  
152 1), 126.9 (C-8), 124.5 (C-6), 115.3 (C-5), 109.9 (C-2),  
153 56.4 (3-OCH<sub>3</sub>).

### 154 **3-Hydroxy-1-(4'-hydroxy-3'**

155 **methoxyphenyl)propan-1-one (4)**: White crystals.  
156 <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta_H$  7.55 (2H, m, H-2',  
157 H-6'), 6.96 (1H, d, 8.0 Hz, H-5'), 4.02 (1H, t, 5.3  
158 Hz, H-3), 3.96 (3H, s, 3'-OCH<sub>3</sub>), 3.18 (1H, t, 5.3  
159 Hz, H-2); <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 125 MHz):  $\delta_C$  199.2  
160 (C-1), 151.0 (C-4'), 146.9 (C-3'), 129.9 (C-1'), 123.8  
161 (C-6'), 114.1 (C-5'), 109.8 (C-2'), 58.5 (C-3), 56.3  
162 (3'-OCH<sub>3</sub>), 40.0 (C-2).

163 **p -Hydroxybenzaldehyde (5)**: white crystals. <sup>1</sup>H-  
164 NMR (acetone-*d*<sub>6</sub>, 500 MHz):  $\delta_H$  9.85 (1H, s, H-  
165 7), 7.80 (2H, d, 8.5 Hz, H-2, H-6), 7.00 (2H, d, 8.5  
166 Hz, H-3, H-5); <sup>13</sup>C-NMR (acetone-*d*<sub>6</sub>, 125 MHz):  $\delta_C$   
167 191.0 (C-7), 164.0 (C-4), 132.8 (C-2, C-6), 130.5 (C-  
168 1), 116.7 (C-3, C-5).

169 **Vanillin (6)**: white needles. <sup>1</sup>H-NMR (methanol-*d*<sub>4</sub>,  
170 500 MHz):  $\delta_H$  9.74 (1H, s, H-7), 7.44-7.41 (2H, m,  
171 H-2, H-6), 6.93 (1H, d, 6.6 Hz, H-5), 3.91 (3H, s,  
172 3-OCH<sub>3</sub>); <sup>13</sup>C-NMR (methanol-*d*<sub>4</sub>, 125 MHz):  $\delta_C$   
173 191.5 (C-7), 153.4 (C-3), 148.3 (C-4), 129.3 (C-6),  
174 126.5 (C-1), 114.9 (C-5), 109.9 (C-2), 55.0 (3-OCH<sub>3</sub>).

175 Cytotoxicity of compounds **1-3** against HepG2 cells  
176 Palmarumycin JC2 (**1**): IC<sub>50</sub> = 36.09 ± 3.79 μg/ml;  
177 5-Hydroxy-4',7-dimethoxyflavone (**2**): IC<sub>50</sub> > 256  
178 μg/ml; Coniferaldehyde (**3**): IC<sub>50</sub> = 53.20 ± 4.37  
179 μg/ml; Ellipticine (positive control): IC<sub>50</sub> = 0.45 ±  
180 0.04 μg/ml.

## 181 DISCUSSION

182 Compound **1** was obtained as yellow needles,  
183  $[\alpha]_D^{25} + 142.4$  ( $c = 0.7$ , CHCl<sub>3</sub>). Its molecular formula  
184 was determined to be C<sub>20</sub>H<sub>14</sub>O<sub>5</sub> by HR-ESI-MS  
185 ([M-H]<sup>-</sup>  $m/z$  333.0758, calculated for C<sub>20</sub>H<sub>13</sub>O<sub>5</sub>  
186 333.0763), indicating that **1** had fourteen degrees of  
187 unsaturation. The <sup>1</sup>H-NMR spectrum showed reso-  
188 nances for the presence of a chelated hydroxy proton  
189 [ $\delta_H$  13.84 (s, 9-OH)], a 1,8-disubstituted naphthalene  
190 moiety [ $\delta_H$  7.55 (2H, m, H-1', H-9'), 7.49 (1H, t,  
191 8.0 Hz, H-2'), 7.44 (1H, t, 7.9 Hz, H-8'), 7.08 (1H,  
192 d, 8.3 Hz, H-3'), 6.92 (1H, d, 7.5 Hz, H-7')], three  
193 aromatic protons of a 1,2,3-trisubstituted benzene  
194 ring [ $\delta_H$  7.55 (1H, m, H-7), 7.33 (1H, dd, 7.6, 1.0

Hz, H-6), 7.08 (1H, d, 8.6 Hz, H-8)], an oxymethine  
195 proton of a secondary alcohol [ $\delta_H$  4.60 (1H, t, 3.7  
196 Hz, H-3), two protons of a methylene group lying  
197 between two electron-withdrawing groups [ $\delta_H$  3.24  
198 (1H, dd, 17.8, 3.4 Hz, H-2a), 2.95 (1H, dd, 17.8, 4.0  
199 Hz, H-2b)]. The <sup>13</sup>C-NMR spectrum had signals  
200 due to 20 carbons, comprising a 1,8-disubstituted  
201 naphthalene moiety carrying two oxygenated car-  
202 bons [ $\delta_C$  147.3 (C-6'), 146.5 (C-4')], a trisubstituted  
203 benzene ring carrying an oxygenated carbon [ $\delta_C$   
204 162.3 (C-9)], a conjugated carbonyl carbon [ $\delta_C$   
205 201.1 (C-1)], a spiroketal carbon [ $\delta_C$  98.9 (C-4)], an  
206 oxymethine carbon [ $\delta_C$  67.4 (C-3)], and a methylene  
207 group [ $\delta_C$  41.4 (C-2)]. The planar structure of **1**  
208 was elucidated by analysis of its HSQC, HMBC, and  
209 COSY spectra. In the COSY spectrum, correlations  
210 between H<sub>2</sub>-2 and H-3 revealed a bond between  
211 C-2 and C-3. In the HMBC spectrum, proton H-2  
212 correlated with C-1, C-3, C-4 and C-10, whereas  
213 proton H-3 presented cross-peaks at C-1, C-4 and  
214 C-5, indicating a 1-tetralone subunit that carried a  
215 secondary alcohol at C-2 and a spiroketal carbon at  
216 C-4. Other HMBC and COSY correlations revealed  
217 the presence of a 1,2,3-trisubstituted benzene ring  
218 of the 1-tetralone subunit and a 1,8-dioxygenated  
219 naphthalene moiety in the molecule (Figure 2). The  
220 spectral data, along with the lack of one degree of  
221 unsaturation, indicated that the 1-tetralone fragment  
222 was bonded to the 1,8-dioxygenated naphthalene  
223 moiety via the spiroketal carbon [ $\delta_C$  98.9 (C-4)]. On  
224 the basis of the spectral analysis and comparison of  
225 the data with the published article,<sup>15,16</sup> compound  
226 **1** could be (3*S*)- almarumycin JC2 ( $[\alpha]_D^{25} + 131.9$  ( $c =$   
227 0.5, CHCl<sub>3</sub>)<sup>15</sup> or the enantiomer (3*R*)- almarumycin  
228 BG1 ( $[\alpha]_D^{25} - 151.0$  ( $c = 0.5$ , CHCl<sub>3</sub>)<sup>16</sup>. Since the  
229 optical rotation of compound **1** was  $[\alpha]_D^{25} + 142.4$   
230 ( $c = 0.7$ , CHCl<sub>3</sub>), it was identified as almarumycin  
231 JC2, which was previously isolated from *Jatropha*  
232 *curcas*.<sup>15</sup>

233 Compound **2** was isolated as a yellow needle. The  
234 <sup>1</sup>H-NMR spectrum showed resonances for the pres-  
235 ence of a chelated hydroxy proton [ $\delta_H$  12.80 (1H,  
236 s, 5-OH)], an isolated olefinic proton [ $\delta_H$  6.57 (s,  
237 H-3)], two *meta*-coupled protons [ $\delta_H$  6.48 (1H, *d*,  
238 2.2 Hz, H-8) and 6.26 (1H, *d*, 2.2 Hz, H-6)], four  
239 aromatic protons of a 1,4-disubstituted benzene ring  
240 [ $\delta_H$  7.84 (2H, *m*, H-2' and H-6') and 7.01 (2H, *m*,  
241 H-3' and H-5')] and two methoxy groups [ $\delta_H$  3.89  
242 and 3.88 (3H each, s, 7-OCH<sub>3</sub> and 4'-OCH<sub>3</sub>)]. The  
243 <sup>13</sup>C-NMR spectrum revealed 17 carbon signals corre-  
244 sponding to a conjugated carbonyl carbon [ $\delta_C$  182.6  
245 (C-4)], fourteen olefinic/aromatic carbons with five  
246 oxygenated carbons ( $\delta_C$  165.6, 164.2, 162.8, 162.4,  
247

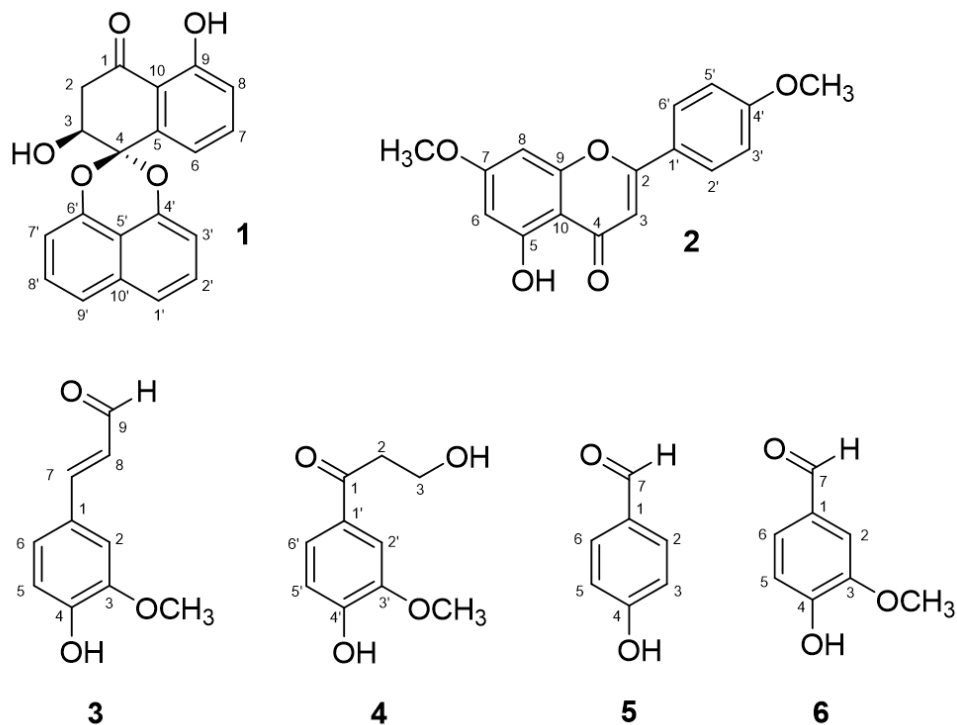


Figure 1: Structures of compounds 1-6

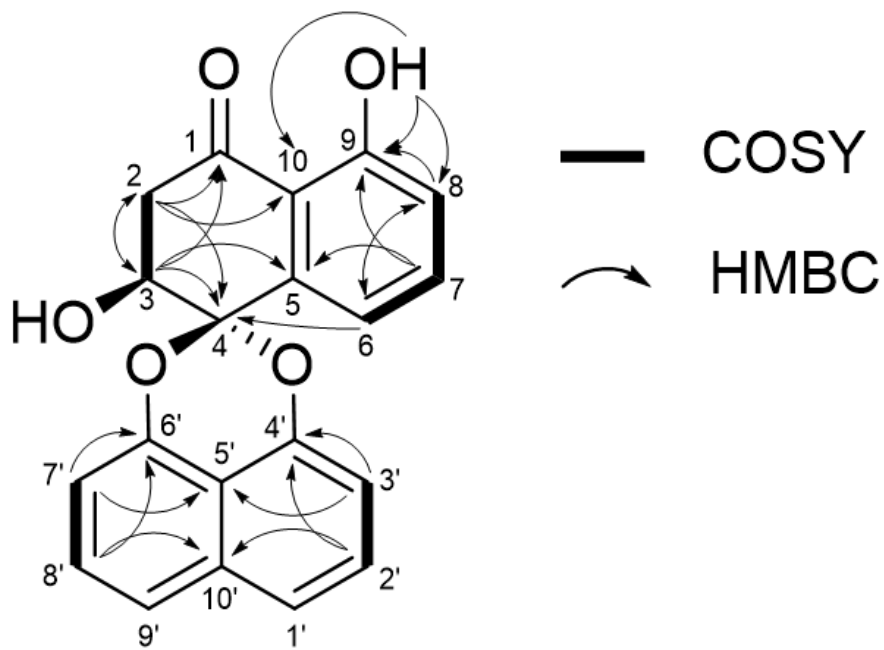


Figure 2:  $^1\text{H}$ - $^1\text{H}$  COSY and selected HMBC correlations of 1

157.9), seven protonated carbons ( $\delta_C$  128.2, 114.7, 104.5, 98.2, 92.8), two fully substituted carbons ( $\delta_C$  123.8, 105.8) and two methoxy groups [ $\delta_C$  55.9 and 55.7 (7-OCH<sub>3</sub> and 4'-OCH<sub>3</sub>)]. The spectral data suggested a flavone carrying a chelated hydroxy group at C-5 and two methoxy groups attached to C-7 and C-4'. A comparison of the NMR data of **2** with those of the reference<sup>17</sup> suggested that **2** was 5-hydroxy-4',7-dimethoxyflavone, which was previously reported in *Combretum zeyheri*.

Compound **3** was obtained as yellow needles. The <sup>1</sup>H-NMR spectrum had resonances for an aldehyde proton [ $\delta_H$  9.65 (1H, d, 7.8 Hz, H-9)], two trans-coupled protons [ $\delta_H$  7.40 (1H, d, 15.8 Hz, H-7), 6.60 (1H, dd, 15.8, 7.8 Hz, H-8)], three aromatic protons of a 1,2,4-trisubstituted benzene ring [ $\delta_H$  7.12 (1H, dd, 8.2, 1.9 Hz, H-6), 7.07 (1H, d, 1.9 Hz, H-2), 6.96 (1H, d, 8.2 Hz, H-5)], and a methoxy group ( $\delta_H$  3.95 (3H, s, 3-OCH<sub>3</sub>)). The <sup>13</sup>C-NMR spectrum showed resonances for 10 carbons, comprising an aldehyde carbonyl carbon [ $\delta_C$  193.9 (C-9)], two olefinic carbons [ $\delta_C$  153.4 (C-7), 126.9 (C-8)], a benzene ring and a methoxy group [ $\delta_C$  56.4 (3-OCH<sub>3</sub>)]. The benzene ring consists of three protonated carbons [ $\delta_C$  124.5 (C-6), 115.4 (C-5), 109.9 (C-2)] and three substituted carbons, two of which are oxygenated [ $\delta_C$  149.3 (C-3), 147.4 (C-4), 127.1 (C-1)]. The above spectral data revealed that the compound had a benzene ring with a conjugated aldehyde, a methoxy group and a hydroxy group. The compound was determined to be 4-hydroxy-3-methoxycinnamaldehyde or coniferaldehyde (**3**) by comparison of the NMR spectral data with those published in the literature<sup>18</sup>.

Compound **4** was isolated as white crystals. The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra closely resembled those of **3**. There were resonances for a 1,3,4-trisubstituted benzene ring carrying two oxygenated carbons [ $\delta_H$  7.55 (2H, m, H-2' and H-6'), 6.96 (1H, d, 8.0 Hz, H-5');  $\delta_C$  151.0 (C-4'), 146.9 (C-3'), 129.9 (C-1'), 123.8 (C-6'), 114.1 (C-5'), 109.8 (C-2')], and a methoxy group [ $\delta_H$  3.96 (3H, s, 3'-OCH<sub>3</sub>);  $\delta_C$  56.3 (3'-OCH<sub>3</sub>)]. The three carbons of the conjugated aldehyde in **3** were replaced by a carbonyl carbon of a ketone [ $\delta_C$  199.2 (C-1)], an oxymethylene group [ $\delta_H$  4.02 (1H, t, 5.3 Hz, H-3);  $\delta_C$  50.5 (C-3)] and a methylene group [ $\delta_H$  3.18 (1H, t, 5.3 Hz, H-2)];  $\delta_C$  40.0 (C-2)]. The spectral data were consistent with those previously reported<sup>19</sup>, suggesting that compound **4** was 3-hydroxy-1-(4'-hydroxy-3'-methoxyphenyl)propan-1-one or  $\beta$ -hydroxypropiovanillone, which was previously isolated from *Cassia laevigata*.

Compound **5** was obtained as white crystals. The <sup>1</sup>H-NMR spectrum had resonances for the aldehyde proton [ $\delta_H$  9.85 (1H, s, H-7)] and four aromatic protons of a 1,4-trisubstituted benzene ring [ $\delta_H$  7.80 (2H, d, 8.5 Hz, H-2 and H-6), 7.00 (2H, d, 8.5 Hz, H-3 and H-5)]. The <sup>13</sup>C-NMR spectrum showed resonances for a conjugated carbonyl carbon [ $\delta_C$  191.0 (C-7)], six aromatic carbons consisting of two pairs of protonated symmetrical carbons [ $\delta_C$  132.8 (C-2, C-6), 116.7 (C-3, C-5)], and two substituted carbons, one of which were oxygenated [ $\delta_C$  164.0 (C-4)]. The compound was therefore determined to be *p*-hydroxybenzaldehyde (**5**)<sup>20</sup>.

Compound **6** was isolated as white needles. The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of **3** were very similar to those of compound **3**. There were signals for an aldehyde [ $\delta_H$  9.74 (1H, s, H-7);  $\delta_C$  191.5 (C-7)], a 1,3,4-trisubstituted benzene ring carrying two oxygenated carbons [ $\delta_H$  7.43-7.41 (2H, m, H-2, H-6), 7.04 (1H, d, 6.6 Hz, H-5);  $\delta_C$  153.4 (C-3), 148.3 (C-4), 129.3 (C-6), 126.5 (C-1), 114.9 (C-5), 109.9 (C-2)], and a methoxy group [ $\delta_H$  3.91 (3H, s, 3-OCH<sub>3</sub>);  $\delta_C$  55.0 (3-OCH<sub>3</sub>)]. The only difference was that the signals for a carbon-carbon double bond disappeared. A comparison of the NMR data with those in the literature<sup>21</sup> suggested that compound **6** was vanillin (**6**).

Compounds **1-3** were evaluated for their in vitro cytotoxicity against HepG2 cells via the MTT assay, with ellipticine used as the positive control. The two polyphenol derivatives, palmarumycin JC2 (**1**) and coniferaldehyde (**3**), had very weak effects, with IC<sub>50</sub> values of 36.09 and 53.20  $\mu$ g/ml, respectively (ellipticine, IC<sub>50</sub> = 0.45  $\mu$ g/ml), whereas 5-hydroxy-4',7-dimethoxyflavone (**2**) was inactive (IC<sub>50</sub> > 256  $\mu$ g/ml). Previously, palmarumycin JC2 (**1**) was reported to exhibit weak or no activity against the NCI-H187, BC, KB, and Vero cell lines<sup>22</sup>. The low cytotoxicity of the compound is consistent with our findings. This is the first report on the cytotoxicity of compounds **2** and **3**.

## CONCLUSION

Six phenolic compounds, palmarumycin JC2 (**1**), 5-hydroxy-4',7-dimethoxyflavone (**2**), coniferaldehyde (**3**), 3-hydroxy-1-(4'-hydroxy-3'-methoxyphenyl)propan-1-one (**4**), *p*-hydroxybenzaldehyde (**5**) and vanillin (**6**), were isolated from the ethyl acetate of the bark of *A. lawii*. This is the first time that six compounds have been found in *A. lawii*. The in vitro cytotoxicity of compounds **1-3** toward HepG2 cells was evaluated via the MTT assay. Nevertheless, the compounds exhibited very weak effects or no activity.

**Table 1: Cytotoxicity of compounds 1-3 against HepG2 cells**

Compound	IC <sub>50</sub> (μg/ml)	IC <sub>50</sub> (μM)
Palmarumycin JC2 (1)	36.09 ± 3.79	108.03 ± 11.35
5-Hydroxy-4',7'-dimethoxyflavone (2)	> 256	> 859
Coniferaldehyde (3)	53.20 ± 4.37	298.88 ± 24.55
Ellipticine <sup>a</sup>	0.45 ± 0.04	1.83 ± 0.16

IC<sub>50</sub> values are expressed as the mean ± standard deviation (n=3).

<sup>a</sup> Positive control

## 351 ABBREVIATIONS

- 352 NMR Nuclear magnetic resonance  
 353 CC Column Chromatography  
 354 COSY Correlation Spectroscopy  
 355 *d* Doublet  
 356 *dd* Doublet of doublet  
 357 GPC Gel Permeation Chromatography  
 358 HMBC Heteronuclear Multiple Bond Coherence  
 359 HSQC Heteronuclear Single Quantum Correlation  
 360 HR-ESI-MS High Resolution Electron Spray Ionization Mass Spectroscopy  
 361  
 362 IC<sub>50</sub> The Half maximal inhibitory concentration  
 363 IR Infrared  
 364 *J* Coupling Constant  
 365 *m* Multiplet  
 366 *q* Quartet  
 367 RP Reversed-phase  
 368 *s* Singlet  
 369 *t* Triplet  
 370 TLC Thin Layer Chromatography

## 371 COMPETING INTEREST

372 The authors declare that they have no conflicts of interest.  
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## 374 AUTHORS' CONTRIBUTION

375 Pham Hoang Quan, Ngo Trang Nhu Ngoc, Nguyen Dieu Lien Hoa: research ideas and project plans;  
 376 Pham Hoang Quan, Nguyen Thi Thao Ly, Trinh Thi Dieu Binh, and Ngo Trang Nhu Ngoc: sample collection, extraction, isolation; structure elucidation;  
 377 Pham Hoang Quan, Nguyen Dieu Lien Hoa: writing the article.  
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